Diagnosis: *Figurata* appears to have no close ally in *Thalassodes* and is distinctively different in wing pattern alone from every other member of the genus.

Distribution: Known from Nandarivatu, Fiji, over 1100 m. above sea level. There is one specimen from Vunidawa, Fiji (R. H. Phillips), in coll. B.M. (N.H.) which was given the MS name of *figurata* by L. B. Prout.

Holotype; ♂ Nandarivatu, 28.vi.1968 at M.V. (H. S. Robinson). In B.M. (N.H.).

Paratypes: 13 ♂♂, 10 ♀♀ Nandarivatu, ix.1967 and 28.vi.1968 at M.V. (H. S. Robinson). In coll. H. S. and G. S. Robinson and in B.M. (N.H.).

I am very grateful to Messrs. D. S. Fletcher and A. H. Hayes of the British Museum (Natural History) and to Dr. L. Davies and Mr. J. Richardson of the Department of Zoology, University of Durham, for their help and encouragement in the preparation of this paper.

Rearing the Jersey Tiger (*Eupalagia quadripunctaria* Poda)

By L. G. F. Waddington

Previous articles of mine on the above subject in issues of Sept. 1963 and Nov. 1964 described how a measure of success finally rewarded my efforts, but two factors still remained to be solved to my satisfaction.

Firstly, the best method of dealing with newly hatched larvae, and secondly the best foodplant.

Thanks to the kindness of my friend at Plympton, I was supplied with batches of ova in August 1965 and August 1967. Only a limited number were retained and the remainder were given to other collectors.

As I was already critical of starting the larvae off in plastic boxes, I decided to use plastic tumblers instead.

I had a supply of these—fitted with bakelite lids—which originally contained Abbey Cane Syrup, marketed by Martineaus of London, and with the aid of a large red hot nail I melted a circle in the lid and an $\frac{1}{8}$" hole in the bottom.

After glueing a circle of muslin inside the lid, they were ready for use; I treated two of these tumblers and stood them in glass tumblers which provided stability and left room for a little water at the bottom for the foodplant.

Dealing with the 1965 batch, I put dead nettle in one tumbler and stinging nettle in the other, making sure that the top of the spray impinged on the muslin.

As soon as the larvae hatched out, they were put straight in the tumblers and these were housed in the garage; it was noticeable that the larvae soon sought the muslin-covered lid where air and food were available.

Condensation was very slight and confined to the bottom half of the tumbler, and consequently no losses occurred through this.

By Oct. 21st many had changed skins 3 times, and on account of the cold were brought into the kitchen.

By Nov. 5th it was obvious that the larvae on dead nettle were much further advanced than those on stinging nettle, so the latter was discontinued; however, it must be stated that in both cases growth was irregular.

The stinging nettle larvae did not take kindly to the change of diet, and many subsequently died.

As the larvae got too large for the tumblers, they were transferred to hurricane glass cages, and by the end of November many were ¾ grown.

Early in December the largest were put into the breeding cage, and some began to spin up in the moss by the middle of the month.

By March 9 the last larva had spun up.

Moths commenced to emerge on Jan. 29 1966 and continued throughout February and March, the last emergence taking place on April 14.

Dealing with the 1967 batch of ova, the same procedure was adopted, except that dead nettle was used exclusively.

On Oct. 1 I put about 20 larvae in a hurricane glass cage and brought it into the kitchen, but after three weeks they showed no more progress than those in the garage; however, on account of the cold in early November, the remainder were brought into the kitchen.

By the end of November a number of larvae were full fed and transferred to the breeding cage.

Bitter cold winds and heavy snowfalls were the order of the day in early December, which ruined the dead nettle and I had recourse to the old sheet-anchor-lettuce, and this was used up to the end.

The first emergence took place on Jan. 18, 1968 and continued during February, the last one taking place on the 22nd; all regrettably typical.

The upshot of these experiments finally convinced me that ventilation is an all important factor in rearing the larvae, particularly in the very early stages, while dead nettle is far and away the best foodplant.

If this is not readily available, I suggest you remove to a locality where it is.

A final thought—the modified methods adopted clearly speeded up larval growth, resulting in emergencies two months earlier than previously experienced.

Current Notes

I am informed of the floating of The British Butterfly Conservation Society, whose title speaks for itself. The problem is a difficult one, and the society's preliminary programme contemplated between 1968 and 1971 is set out as follows:

(a) Assisting in an evaluation of the present status of species in the wild;
(b) examining, assessing and reporting of potentialities regarding particular species requiring conservation;
(c) general discussions with statutory and voluntary bodies enabled to assist in conservaton;
(d) specific discussions with specific bodies on possible projects for conservation in the wild, and breeding and re-introductions.

Peter Scott is named as President, Thomas Frankland as Chairman, and Robert Goodden, of Over Compton, Sherborne, Dorset, as secretary. —Ed.